MedPharmRes

Journal of University of Medicine and Pharmacy at Ho Chi Minh City homepage: http://www.medpharmres.vn/ and http://www.medpharmres.com/

Original article

Pheochromocytoma: Impact of genetic testing on clinical practice in Vietnam

Minh Duc Do^{a#}, Thang Viet Tran^{b#}, Hoang Linh Le Gia^a, Hoang Van Lam^c, Hen Huu Phan^c, Minh Binh Ta^c, An Thuy Thi Nguyen^c, Ngoc The Phung^d, Diana E Benn^e, Bruce G Robinson^e, Vu Anh Hoang^a, Thao Phuong Mai^{f*}

^aCenter for Molecular Biomedicine, University of Medicine and Pharmacy at Ho Chi Minh City, Vietnam;

^bDepartment of Endocrinology, Faculty of Medicine, University of Medicine and Pharmacy at Ho Chi Minh City, Vietnam; ^cDepartment of Endocrinology, Cho Ray Hospital, Ho Chi Minh City, Vietnam;

^dDepartment of Endocrinology, People 115 Hospital, Ho Chi Minh City, Vietnam;

^eCancer Genetics, Kolling Institute, Royal North Shore Hospital & University of Sydney, St Leonards, New South Wales, 2065, Australia;

^fDepartment of Physiology – Pathophysiology – Immunology, Faculty of Medicine, University of Medicine and Pharmacy at Ho Chi Minh City, Vietnam.

Received November 21, 2020: Revised February 05, 2021: Accepted February 09, 2021

Abstract: *Introduction:* Germline mutations in predisposing genes have been found in 30-40% of pheochromocytoma/paraganglioma patients. Screening for inherited genetic mutations provide clinicians with mutation-positive patient management strategies in addition to identifying family members at risk of disease. However, genetic testing for pheochromocytoma has not been performed widely in Vietnam. *Methods:* Seven patients diagnosed with pheochromocytoma in Vietnam underwent germline genetic testing in known pheochromocytoma-associated genes by direct sequencing. When a germline mutation was identified the first-degree relatives were counseled and offered genetic testing for the inherited mutation. *Results:* Mutations were found in five of seven cases and all mutations were in *RET* proto-oncogene codon 634 indicating a high risk of developing aggressive medullary thyroid cancer and in some cases leading to prophylactic thyroidectomy as recommended. *Conclusions:* Genetic testing plays an essential role in the clinical management of pheochromocytoma patients. Genetic results have significantly changed the clinical approach in these patients and identified 'at risk' family members.

Keywords: genetic testing; germline mutation; pheochromocytoma.

1. INTRODUCTION

Pheochromocytomas are tumors arising from the catecholamine-producing cells in the adrenal medulla. The typical symptoms of pheochromocytoma include headache, sweating, tremor, pallor and palpitations. Due to excessive

The two authors have equal contribution DOI: 10.32895/UMP.MPR.5.2.3

paroxysmal catecholamine secretion into the bloodstream, the symptoms are usually episodic and associated with considerable cardiovascular complications and even death [1-3]. Previously, only 10% of pheochromocytoma cases were thought to be hereditary as proposed by Emanuel Bravo in 1984 [4]. Recently, with the development of genetic

© 2021 MedPharmRes





^{*}Address correspondence to Thao Phuong Mai at the Department of Physiology – Pathophysiology – Immunology, Faculty of Medicine, University of Medicine and Pharmacy at Ho Chi Minh City, Vietnam; E-mail: <u>drmaithao@ump.edu.vn</u>

sequencing technologies, germline mutations have been found to contribute up to 30–40% of the pheochromocytoma (and paraganglioma) cases [5-8]. Genetic testing for all cases of pheochromocytoma/paraganglioma are now recommended as part of the standard care [6, 9-11]. In addition to guiding patient management, genetic results also provide clinicians with the ability to screen for pheochromocytoma and related tumors in relatives of a mutation-positive proband [12-14]. However, genetic testing is not routinely performed in Vietnam due to the limitation of technologies and resources. Here we report the first pheochromocytoma cases that were referred to our center for genetic testing and how the results have impacted clinical practice.

2. MATERIALS AND METHOD

2.1. Subjects

This study was approved by the Ethical Committee of The University of Medicine and Pharmacy at Ho Chi Minh City, Vietnam. Patients were admitted to Cho Ray Hospital and University Medical Center between 2016 and 2018. Pheochromocytomas were diagnosed based on the elevation of 24-hour urinary catecholamine and plasma free metanephrine, computed tomography (CT) imaging, and postoperative pathology. Other clinical information including age, gender, syndromic characteristics of multiple endocrine neoplasia type 2, neurofibromatosis type 1, Von Hippel-Lindau disease, familial history of pheochromocytoma, medullary thyroid carcinoma, head and neck paraganglioma, serum calcium, parathyroid hormone, and calcitonin were also recorded. Patients were counseled and provided written informed consent for genetic testing. If a germline mutation was found, all first-degree relatives of mutation-positive patients were also counseled and provided written informed consent for genetic testing. If genetic testing was indicated in the children, the written informed consent was obtained from their next of kin, caretakers, or guardians.

2.2. Biochemical tests

All the results of biochemical diagnostic tests in this study were performed in the standard biochemical laboratory of either Cho Ray hospital or the University Medical Center following the criteria of ISO 15189:2012.

2.3. DNA extraction

GeneJET Whole Blood Genomic DNA Purification Mini Kit (Thermo Fisher Scientific, Massachusetts, USA) was used to extract the genomic DNA of the subjects according to the manufacturer's protocol and these samples were stored at -20°C for further experiments.

2.4. Genetic testing

Genetic testing for pheochromocytoma patients was performed following the American Endocrine Society decisional algorithm [6]. All coding exons and flanking regions of VHL (GenBank NG_008212.3), SDHB (GenBank NG_012340.1), SDHC (GenBank NG_012767.1), SDHD (GenBank NG_012337.3), MAX (GenBank NG_029830.1), TMEM127 (GenBank NG_027695.1) genes and RET (GenBank NG_007489.1) exons 10, 11, 13, 14, 15, 16 were amplified and sequenced. DNA amplification was performed by Mastercycler@proS, the results of DNA electrophoresis on 1% agarose gels with Diamond[™] Nucleic Acid Dye (Promega, Madison, WI, USA) were used to confirm the appropriate length of amplified products. These products were subsequently purified with Exosap-IT glycerol solution (Thermo Fisher Scientific, Massachusetts, USA) and then sequenced by BigDye® Terminator v3.1 Cycle Sequencing kit (Applied Biosystems, Foster City, CA, USA). Sequencing reactions were analyzed with ABI 3130 Genetic Analyzer (Applied Biosystems). Results were compared to the reference sequences of target genes in Genebank. All the primers for genetic testing are listed in Supplementary Table 1.

3. RESULTS

All patients (4 females and 3 males) referred to our center were at high risk of germline mutation as they were diagnosed with bilateral pheochromocytoma and/or at early age of onset, and/or had a familial history of either pheochromocytoma or medullary thyroid cancer. Two patients BN01 (with unilateral pheochromocytoma) and BN02 (with bilateral pheochromocytoma) were tested for VHL, SDHB, SDHC, SDHD, MAX, TMEM127, and RET mutations but none were detected. Multiple endocrine neoplasia type 2A (MEN2A) manifestations were found in 3 out of 7 cases (BN05, BN06, BN07), the other patients were diagnosed with pheochromocytoma only. None of the probands showed manifestations or family history of Neurofibromatosis type 1, von-Hippel-Lindau syndrome or paraganglioma. Four of seven patients (BN02, BN03, BN04, and BN07) were diagnosed with bilateral pheochromocytoma, whilst BN05 and BN06 developed another pheochromocytoma after the first adrenalectomy surgery. Clinical characteristics of these patients are summarized in Table 1. Family history of these patients also showed that BN03, BN04 and BN05 had siblings who developed either pheochromocytoma or medullary thyroid cancer and the deaths of these family members were described as sudden and at a very young age (from 30s to 40s years old).

In patients with MEN2A syndromic manifestations, RET gene sequencing was indicated. On the other hand, the order of genetic testing in patients with only signs and symptoms of pheochromocytoma were performed according to the Endocrine Society decisional algorithm6. RET gene mutations were found in 3 out of 3 MEN2A patients (BN05, BN06, and BN07) and in two of three patients with bilateral pheochromocytoma (BN03 and BN04). All RET mutations were in codon 634 and either c.1900T>C or c.1900T>G (Table 1).

Genetic analysis in the first degree relatives of mutationpositive probands showed an inherited pattern in all the families except the family of BN05 and BN06. However, patient BN05 presented with typical MEN2A phenotype and medullary thyroid cancer was reported in one of his siblings (who declined genetic testing) suggesting that the disease in this family was highly likely inherited. Interestingly, genetic testing in patient BN06 suggested that this patient carried a 'de novo' mutation as none of the first degree family members carried this RET mutation. Currently, the screening for disease has been undertaken periodically for mutation carriers without any phenotypic signs and symptoms. The pedigrees of mutation-positive probands and their relatives' mutation status are illustrated in Figure 1.

	Age at diagnosis/ Gender	Adrenal tumor (size on CT imaging)	Syndromic manifestati ons	hromocytoma patients 24h-Urinary catecholamine (µg/24h)			Plasma metanep hrine (<90pg/ mL)	Calciton in (<6.4pg/ mL)	CEA (<5ng/ mL)	Calciu m (mmol /L)	iPTH (21- 45pg/ mL)	Genetic mutation
				Adren alin (<20)	Noradre nalin (<90)	Dopa mine (<600)						
BN 01	30/F	Unilateral L: 30x40x50 mm	No	57.9	340.7	430.5	64.5	5.1	3.2	2.48	33.9	ND
BN 02	32/M	Bilateral L: 20x30x35 mm R: 40x45x40 mm	No	13.1	250.4	500.4	102.5	4.2	2.2	2.52	20.1	ND
BN 03	33/F	Bilateral L: 40x50x60 mm R: 50x40x55 mm	No	7.6	199	278.3	758	4.1	3.8	2.30	29.8	RET gene: c.1900T >C (Cys634 Arg)
BN 04	34/M	Bilateral L: 15x20x20 mm R: 25x40x30 mm	No	133.3	116.4	568.4	589.8	57.2	14.6	2.42	25.4	<i>RET</i> gene: c.1900T >G (Cys634 Gly)
BN 05	41/M	Bilateral L: 12x10x10 mm* R: 60x90x70 mm	MEN2A (MTC & PC)	460.0	1098.6	1228.4	1102.0	869.0	85.5	2.34	33.6	<i>RET</i> gene: c.1900T >C (Cys634 Arg)
BN 06	25/F	Bilateral L: 59x55x40 mm R: 36x48x25 mm*	MEN2A (MTC & PC)	76.2	210	462.0	861.3	28.6	33.5	2.20	42.5	RET gene: c.1900T >C (Cys634 Arg)
BN 07	30/F	Bilateral L: 15x20x20 mm R: 25x40x30 mm	MEN2A (MTC & Bilateral PC)	800	311	1400.0	717.0	2069	51.8	2.32	46	RET gene: c.1900T >C (Cys634 Arg)

Table 1. Clinical characteristics of pheochromocytoma patients

M, Male; F, Female

L, left pheochromocytoma tumor; R, right pheochromocytoma tumor

ND, not detectable

PC, Pheochromocytoma

MTC, Medullary thyroid carcinoma

*, First PC diagnosis followed by subsequent PC diagnosis in contralateral gland.

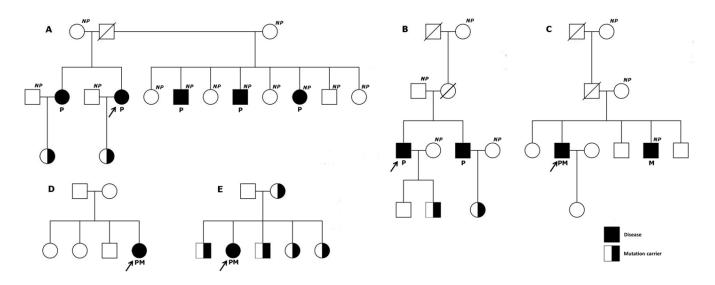


Figure 1. Pedigree of pheochromocytoma patients who carried germline mutation

Arrow indicates the patients in this study. Objects were classified as Disease if they were diagnosed with either pheochromocytoma, medullary thyroid carcinoma or both; objects were classified as Mutation carrier if they carry pathogenic mutation without detectable phenotype at the time of examination. P, Pheochromocytoma; M, Medullary thyroid carcinoma; NP, Genetic testing was not performed; A: Pedigree of patient BN03; B: Pedigree of patient BN04; C: Pedigree of patient BN05; D: Pedigree of patient BN06; E: Pedigree of patient BN07

4. DISCUSSION

Despite the fact that germline mutations may account for up to a total of 30-40% pheochromocytoma/paraganglioma cases and this disease can beheritable [5, 15], genetic testing for pheochromocytoma is still not routinely performed in Vietnamese patients. In this study, we found that five of six patients with bilateral pheochromocytoma carried germline mutation in codon 634 of *RET* proto-oncogene and four of five mutation-positive probands showed increased plasma adrenaline and noradrenaline.

We also reported a potential 'de novo' mutation of RET in patient BN06. It has been reported that 'de novo' RET mutations have been identified in as many as 9% of MEN2A and medullary thyroid carcinoma (MTC) cases and this once again emphasizes the role of genetic testing in patients who are diagnosed with pheochromocytoma/MTC without any apparent family history [16]. It should be noted that up to 7.3% of RET germline mutation carriers have no family history [17, 18]. Knowledge regarding the genetic background of patients with pheochromocytoma has the potential to improve management by providing a more accurate risk stratification and follow-up plan as well as genetic counseling and prophylactic thyroid surgery [14, 19]. For example in the case of patient BN04, after RET mutation identification, the patient was referred for thyroid examination. Thyroid ultrasound described a hypoechoic nodule 4x7 mm and the result of fine needle aspiration was 'normal'. The patient agreed to undergo prophylactic total thyroidectomy and the pathology result of the thyroid nodule showed medullary thyroid cancer.

It is very important to understand the genetic status of firstdegree relatives of mutation-positive patients. Once specific mutations are found in the relatives, it will totally change the medical strategies for screening and monitoring of related tumors¹⁴. In patients BN04 and BN07 family; genetic counseling, monitoring plan, and prophylactic thyroid surgery were offered to all the patients' family members who carried the *RET* mutation. Even if the prophylactic thyroidectomy is not performed, physical examination, ultrasound, and biochemical monotoring annually are very useful for these subjects.

This study has several limitations that need to be discussed. First, the sample size is small due to the fact that pheochromocytoma is not a prevalent disease and only patients with a high risk for having genetic mutation were recruited. This selection bias may overemphasize the benefits of genetic testing. Second, because of the disadvantage of direct sequencing technique, neither *SDHA*, *SDHAF2* or other PC/PGL candidate genes were included in the genetic testing panel. Advanced sequencing technique which was proven competent for long and complex genes can be applied in the future to better detect genetic mutations in pheochromocytoma [20-22].

Current genetic testing algorithms are based either on the 2014 American College of Medical Genetics or the 2014 Endocrine Society Clinical Practice Guidelines [6, 9]. Both guidelines recommend that all patients with pheochromocytoma should consider genetic testing as part of their management. However, to our knowledge, this is the first report of genetic testing in a series of Vietnamese pheochromocytoma patients in Vietnam and we expect that this result will leverage the role of genetic testing of pheochromocytoma patients to overcome barriers such as high cost and social-emotional concern.

Conclusion

Our work highlights the importance of genetic testing of pheochromocytoma patients leading to improved management and early detection of disease in their mutation-positive family members.

LIST OF ABBREVIATIONS

MEN2A: multiple endocrine neoplasia type 2A; MTC: medullary thyroid carcinoma.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

DATA AVAILABLE STATEMENT

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request: please contact drmaithao@ump.edu.vn.

FUNDING

This study was partially funded by the University of Medicine and Pharmacy at Ho Chi Minh City, Vietnam. Funding number 103/2020/HD-DHYD. The Funder has no role on the experiment design, paper writing, and publication.

AUTHORS' CONTRIBUTION

MDD, TVT, TPM designed the study. MDD, TVT, HVL, HHP, MBT, ATTN, NTP included the patients to the study. HLLG, VAH, TPM performed DNA sequencing. MDD, TVT, DEB, BGR, TPM analyzed the data and wrote the manuscript.

ACKNOWLEDGEMENTS

We thank Associate Professor Roderick Clifton-Bligh, Cancer Genetics, Kolling Institute for helpful advice and support.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

This study was approved by the Ethical Committee of University of Medicine and Pharmacy at Ho Chi Minh City, Vietnam. The ID number for ethics approval is 263/HĐĐĐ-ĐHYD. All the patients and their family members have read, fully understood and agreed by signing the informed consent to participate in this study.

CONSENT FOR PUBLICATION

All the patients and their family members fully understood and agreed by signing in the informed consent that their disease status and pedigrees can be published anonymously.

ORCID ID

Minh Duc Do ^{[D}https://orcid.org/0000-0002-9997-6390]

Thao Phuong Mai ^bhttps://orcid.org/0000-0002-6045-099X

REFERENCES

- Gimm O, Koch CA, Januszewicz A, Opocher G, Neumann HP. The genetic basis of pheochromocytoma. *Front Horm Res.* 2004;31:45-60.
- Neumann HPH, Young WF, Eng C. Pheochromocytoma and Paraganglioma. N Engl J Med. 2019;381(6):552-565. doi:10.1056/NEJMra1806651
- Canu L, Parenti G, De Filpo G, Mannelli M. Pheochromocytomas and Paragangliomas as Causes of Endocrine Hypertension. *Front Endocrinol*. 2019;10:333. doi:10.3389/fendo.2019.00333

- Bravo EL, Gifford RW. Current concepts. Pheochromocytoma: diagnosis, localization and management. N Engl J Med. 1984;311(20):1298-1303. doi:10.1056/NEJM198411153112007
- Favier J, Amar L, Gimenez-Roqueplo A-P. Paraganglioma and phaeochromocytoma: from genetics to personalized medicine. *Nat Rev Endocrinol.* 2015;11(2):101-111. doi:10.1038/nrendo.2014.188
- Lenders JWM, Duh Q-Y, Eisenhofer G, et al. Pheochromocytoma and paraganglioma: an endocrine society clinical practice guideline. J Clin Endocrinol Metab. 2014;99(6):1915-1942. doi:10.1210/jc.2014-1498
- Currás-Freixes M, Inglada-Pérez L, Mancikova V, et al. Recommendations for somatic and germline genetic testing of single pheochromocytoma and paraganglioma based on findings from a series of 329 patients. J Med Genet. 2015;52(10):647-656. doi:10.1136/jmedgenet-2015-103218
- Karasek D, Frysak Z, Pacak K. Genetic Testing for Pheochromocytoma. Curr Hypertens Rep. 2010;12(6):456-464. doi:10.1007/s11906-010-0151-1
- 9. Hampel H, Bennett RL, Buchanan A, Pearlman R, Wiesner GL, Guideline Development Group, American College of Medical Genetics and Genomics Professional Practice and Guidelines Committee and National Society of Genetic Counselors Practice Guidelines Committee. A practice guideline from the American College of Medical Genetics and Genomics and the National Society of Genetic Counselors: referral indications for cancer predisposition assessment. *Genet Med Off J Am Coll Med Genet.* 2015;17(1):70-87. doi:10.1038/gim.2014.147
- Gimenez-Roqueplo A-P, Lehnert H, Mannelli M, et al. Phaeochromocytoma, new genes and screening strategies. *Clin Endocrinol* (*Oxf*). 2006;65(6):699-705. doi:10.1111/j.1365-2265.2006.02714.x
- Mercado-Asis LB, Wolf KI, Jochmanova I, Taïeb D. PHEOCHROMOCYTOMA: A GENETIC AND DIAGNOSTIC UPDATE. Endocr Pract Off J Am Coll Endocrinol Am Assoc Clin Endocrinol. 2018;24(1):78-90. doi:10.4158/EP-2017-0057
- Amar L, Baudin E, Burnichon N, et al. Succinate dehydrogenase B gene mutations predict survival in patients with malignant pheochromocytomas or paragangliomas. J Clin Endocrinol Metab. 2007;92(10):3822-3828. doi:10.1210/jc.2007-0709
- Benn DE, Gimenez-Roqueplo A-P, Reilly JR, et al. Clinical presentation and penetrance of pheochromocytoma/paraganglioma syndromes. J Clin Endocrinol Metab. 2006;91(3):827-836. doi:10.1210/jc.2005-1862
- Wells SA, Asa SL, Dralle H, et al. Revised American Thyroid Association guidelines for the management of medullary thyroid carcinoma. *Thyroid Off J Am Thyroid Assoc.* 2015;25(6):567-610. doi:10.1089/thy.2014.0335
- Opocher G, Schiavi F. Genetics of pheochromocytomas and paragangliomas. *Best Pract Res Clin Endocrinol Metab*. 2010;24(6):943-956. doi:10.1016/j.beem.2010.05.001
- 16. Schuffenecker I, Ginet N, Goldgar D, et al. Prevalence and parental origin of de novo RET mutations in multiple endocrine neoplasia type 2A and familial medullary thyroid carcinoma. Le Groupe d'Etude des Tumeurs a Calcitonine. Am J Hum Genet. 1997;60(1):233-237.
- Learoyd DL, Robinson BG. Does genetic screening increase detection of familial medullary thyroid cancer in apparently sporadic cases? *Nat Clin Pract Endocrinol Metab.* 2008;4(3):132-133. doi:10.1038/ncpendmet0722
- Elisei R, Romei C, Cosci B, et al. RET genetic screening in patients with medullary thyroid cancer and their relatives: experience with 807 individuals at one center. *J Clin Endocrinol Metab.* 2007;92(12):4725-4729. doi:10.1210/jc.2007-1005
- Muth A, Crona J, Gimm O, et al. Genetic testing and surveillance guidelines in hereditary pheochromocytoma and paraganglioma. *J Intern Med.* 2019;285(2):187-204. doi:10.1111/joim.12869
- Ben Aim L, Pigny P, Castro-Vega LJ, et al. Targeted next-generation sequencing detects rare genetic events in pheochromocytoma and paraganglioma. J Med Genet. 2019;56(8):513-520. doi:10.1136/jmedgenet-2018-105714
- Nguyen HT, Tran DH, Ngo QD, et al. Evaluation of a Liquid Biopsy Protocol using Ultra-Deep Massive Parallel Sequencing for Detecting and Quantifying Circulation Tumor DNA in Colorectal Cancer Patients. *Cancer Invest*. 2020;38(2):85-93. doi:10.1080/07357907.2020.1713350
- Do MD, Le LGH, Nguyen VT, et al. High-Resolution HLA Typing of HLA-A, -B, -C, -DRB1, and -DQB1 in Kinh Vietnamese by Using Next-Generation Sequencing. *Front Genet*. 2020;11:383. doi:10.3389/fgene.2020.00383